

Screening of Secondary Metabolites and Antibacterial Activity of Some Indian Spices

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ABSTRACT

Current research work deals with screening of secondary metabolites and antibacterial potential of *Rhizoma zingiberis*, *Cuminum cyminum* L., *Curcuma longa* L., *Piper nigrum* L. and *Cinnamomum tamala* the Indian spices. Ethanol and water were used for preparation of test extracts. The present Indian spices found rich in secondary metabolites and shown presence of significant antimicrobial properties. The ethanolic extract of *Curcuma longa*, aqueous extract of *Cuminum cyminum*, *Cinnamomum tamala* and *Piper nigrum* yielded more phytochemicals while for *Rhizoma zingiberis* both the solvents found effective. Ethanolic extract of *Piper nigrum* and *Rhizoma zingiberis* found more effective antibacterial among all the spices. The results obtained may support the use of the spices in traditional medicine for the treatment of various diseases and in drug developments.

Keywords: Indian spices, Secondary metabolites, Antibacterial.

INTRODUCTION

Plants are the effective source of secondary metabolites used in traditional as well as modern medicines and most of the organisms depend on plants for their existence (Patil *et al.*, 2014).

Indian spices include a variety of spices grown across the country and these are used to enhance flavor and aroma of the foods and it also provide antimicrobial properties (Nanasombat *et al.*, 2002). It may also contribute in piquancy of foods and beverages (Praveen and Nazia, 2006). Spices are the most commonly used natural antimicrobial agents

in foods. Some of the natural compounds found in various spices possess antimicrobial activity (Indu *et al.*, 2006) and as the evaluation of any drug is based on phytochemical and pharmacological approaches the present research work was undertaken to screen out the qualitative phytochemical content of the Indian spices like *Rhizoma zingiberis*, *Cuminum cyminum* L., *Curcuma longa* L., *Piper nigrum* L. and *Cinnamomum tamala* using different solvents like water and ethanol along with its antibacterial activity.

MATERIALS AND METHODS

Collection and Authentication:

Fresh and healthy Indian spices like *Rhizoma zingiberis*, *Cuminum cyminum* L., *Curcuma longa* L., *Piper nigrum* L. and *Cinnamomum tamala* were brought from the Local market of Gadhinglaj City, Maharashtra, India during April 2015. All spices were authenticated by Prof. R. S. Sawant, Department of Botany, Dr. Ghali College, Gadhinglaj, Kolhapur district, Maharashtra, India.

Preparation of extracts for Phyto-chemical analysis:

150 gm of dried powder of spices was mixed with 500 ml of ethanol for each. After filtration, the filtrate

How to cite this article:

Ashvin G. Godghate, Rahul Shivaji Patil and Rajaram S. Sawant (2015). Screening of Secondary Metabolites and Antibacterial Activity of Some Indian Spices. *Biolife*, 3(3), pp 614-619. doi:10.17812/blj.2015.337

Published online: 7 July, 2015

was dried and used for phytochemical test. For water extract 150 gm of powder were mixed with 1500 ml of distilled water and heat on water bath for 1/3 rd of original concentration and used for further analysis (Godghate and Sawant, 2014).

Identification tests for Phyto-chemical analysis:

The aqueous and ethanolic extracts of the spices were analyzed for the qualitative phytochemicals analysis as shown in table 1 and 2 using standard methods (Damodaran and Manohar, 2012; Patil and Bhise, 2015; Harborne, 1973; Sawant and Godghate, 2013; ^aPatil *et al*, 2015; Sofowora, 1993).

Preparation of test extracts for antibacterial activity:

The aqueous and ethanolic extracts of spices were prepared by addition of 1 gm of respective powder into 10 ml of respective solvents and kept at room temperature for overnight. Sample further used after its centrifugation.

Test organisms:

The standard test microorganisms used in current study were obtained from National Collection of Industrial Microorganisms, Pune, (M.S.) India are as following: *Bacillus subtilis* NCIM 2635, *Proteus vulgaris* NCIM 2813, *Salmonella typhimurium* NCIM 2501 and *Staphylococcus aureus* NCIM 2654.

Preparation of bacterial suspension:

A loop full suspension of the test organisms were aseptically streaked onto nutrient agar slants and incubated (at 37°C for 24 hours). On next day bacterial growth was harvested from the respective slant and suspension was prepared using sterile 1 ml normal saline. The all suspensions stored in the refrigerator at 4°C until used (^bPatil *et al*, 2015).

Antibacterial activity:

Antibacterial activity of the various spices in aqueous and ethanolic extracts were studied against the various bacteria (Table 3) was determined by using agar well diffusion method on Nutrient agar medium.

Table 1: Phytochemicals of Aqueous (A) and Ethanolic (E) extract of *Rhizoma zingiberis*, *Cuminum cyminum*, and *Curcuma longa*.

Sr. No.	Test		Results					
			<i>Rhizoma zingiberis</i>		<i>Cuminum cyminum</i>		<i>Curcuma longa</i>	
			A	E	A	E	A	E
1	Alkaloids:	Wagner's reagent	+	+	+	-	-	+
		Hager's reagent	+	+	+	+	+	-
2	Saponin: Foam test		+	+	+	+	+	+
3	Tannin: FeCl ₃		-	-	+	-	-	-
4	Steroids		-	-	-	-	+	-
5	Anthocyanin		-	-	-	-	-	+
6	Coumarin		-	+	-	+	-	-
7	Chalcones		-	-	-	-	+	-
8	Protein		-	-	+	+	+	-
9	Amino acids		-	-	-	-	-	-
10	Flavonoids:	Alkaline Reagent Test	+	+	+	+	-	-
		NH ₄ OH	-	+	+	+	-	-
11	Diterpenes		-	+	+	-	-	-
12	Phytosterol		-	-	-	-	-	+
13	Phenols: FeCl ₃ test		-	-	+	-	-	+
14	Phlobatannins		+	-	-	-	-	+
15	Leucoanthocyanin		-	-	-	-	-	-
16	Anthroquinone		+	-	-	-	+	+
17	Emodins		-	-	-	-	+	+
18	Cardial Glycosides: Keller-Killani test		-	-	+	+	-	+
19	Carbohydrates: Barfoed's reagent		-	-	-	-	-	+
20	Acid		-	-	-	-	-	-

RESULTS AND DISCUSSION

Phytochemical analysis of aqueous and ethanolic extracts of the spices was carried out and results are presented in Table 1 and 2.

The aqueous extract of *Rhizoma zingiberis* was found with the phytochemicals like alkaloids, saponin, flavonoids, phlobatannin and anthroquinone; the ethanolic extract shown presence of alkaloids, saponin, coumarin, flavonoids and diterpenes.

Aqueous extract of *Cuminum cyminum* L. shown presence of the secondary metabolites like alkaloids, saponin, tannin, proteins, flavonoids, diterpenes, phenol and cardial glycosides and shown its efficiency more than the ethanolic extract, other hand the ethanolic extract shown presence of alkaloids, saponin, coumarin, protein, flavonoids and cardial glycosides.

The aqueous extract with *Curcuma longa* L. shown presence of alkaloids, saponin, steroids, emodins, proteins, anthroquinone and chalcones

while, the ethanolic extract shown presence of alkaloids, saponin, anthocyanin, emodins, phytosterol, phlobatannin, phenol, anthroquinone, crdial glycosides and carbohydrates and found more efficient.

The aqueous extract of *Piper nigrum* L. yielded the phytochemicals like alkaloids, saponin, steroids, proteins, flavonoids, phytosterol and phlobatannin while, its ethanolic extract contains only few secondary metabolites like alkaloids, proteins and diterpenes.

Aqueous extract of *Cinnamomum tamala* shown presence of phytochemicals like alkaloids, saponin, coumarin and flavonoids while, its ethanolic extract shown presence of alkaloids, saponin and tannin only.

Flavonoids have anti-inflammatory, antimicrobial (Baez et al, 1999; Ogundipe, 2001; Xu HX and Lee, 2001), antioxidant, vascular activities along with other medicinal properties (Harborne and Willians, 1992). Tannin may be toxic to microorganisms like bacteria, yeast and filamentous fungi (Harborne, 1973). It have

Table 2: Phytochemicals of Aqueous (A) and Ethanolic (E) extract of *Piper nigrum* and *Cinnamomum tamala*

Sr. No.	Test		Results			
			<i>Piper nigrum</i>		<i>Cinnamomum tamala</i>	
			A	E	A	E
1	Alkaloids:	Wagner's reagent	+	+	+	-
		Hager's reagent	+	-	+	+
2	Saponin: Foam test		+	-	+	+
3	Tannin: FeCl ₃		-	-	-	+
4	Steroids		+	-	-	-
5	Anthocyanin		-	-	-	-
6	Coumarin		-	-	+	-
7	Chalcones		-	-	-	-
8	Protein		+	+	-	-
9	Amino acids		-	-	-	-
10	Flavonoids:	Alkaline Reagent Test	+	-	+	-
		NH ₄ OH	-	-	+	-
11	Diterpenes		-	+	-	-
12	Phytosterol		+	-	-	-
13	Phenols: FeCl ₃ test		-	-	-	-
14	Phlobatannins		+	-	-	-
15	Leucoanthocyanin		-	-	-	-
16	Anthroquinone		-	-	-	-
17	Emodins		-	-	-	-
18	Cardial Glycosides: Keller-Killani test		-	-	-	-
19	Carbohydrates: Barfoed's reagent		-	-	-	-
20	Acid		-	-	-	-

Key: (+) Positive test, (-) Negative test

Table 3: Antimicrobial activity of Aqueous (A) and Ethanolic (E) extract of *Rhizoma zingiberis* and *Cuminum cyminum*

Organism used	Zone of inhibition (in mm)											
	<i>Rhizoma zingiberis</i> (10 %)						<i>Cuminum cyminum</i> (10 %)					
	A		E		Control		A		E		Control	
	50µl	100µl	50µl	100µl	50µl	100µl	50µl	100µl	50µl	100µl	50µl	100µl
<i>Bacillus subtilis</i> NCIM 2635	-	-	15.0± 1.00	18.6± 0.57	-	-	-	-	-	-	-	-
<i>Proteus vulgaris</i> NCIM 2813	-	-	-	-	-	-	-	16.6± 0.57	18.0 ±1.0 0	22.0± 1.00	-	-
<i>Salmonella typhimurium</i> NCIM 2501	-	-	20.0± 1.00	18.0± 1.00	-	-	-	-	-	-	-	-
<i>Staphylococcus aureus</i> NCIM 2654	-	-	14.6± 0.57	18.0± 1.15	-	-	-	-	-	-	-	-

Note: Each value is the mean of three readings ± SD.

Table 4: Antimicrobial activity of Aqueous (A) and Ethanolic (E) extract of *Curcuma longa* and *Piper nigrum*

Organism used	Zone of inhibition (in mm)											
	<i>Curcuma longa</i> (10 %)						<i>Piper nigrum</i> (10 %)					
	A		E		Control		A		E		Control	
	50µl	100µl	50µl	100µl	50µl	100µl	50µl	100µl	50µl	100µl	50µl	100µl
<i>Bacillus subtilis</i> NCIM 2635	-	-	16.0± 1.00	12.6± 0.57	-	-	-	-	16.0± 1.00	21.0± 1.00	-	-
<i>Proteus vulgaris</i> NCIM 2813	-	-	-	-	-	-	-	-	-	-	-	-
<i>Salmonella typhimurium</i> NCIM 2501	-	-	-	-	-	-	-	-	12.0± 1.00	22.0± 1.00	-	-
<i>Staphylococcus aureus</i> NCIM 2654	-	-	-	-	-	-	-	-	14.0± 1.00	16.0± 1.00	-	-

potential antiviral (Lin *et al*, 2004; Krishna *et al*, 2014) and antibacterial activity (Akiyama *et al*, 2001; Funatogawa *et al*, 2004; Venkanna Lunavath and Estari Mamidala, 2013).

The data (Table 3, 4 and 5) revealed about antibacterial activity of the spices. Only the ethanolic extract of *Rhizoma zingiberis* shown antibacterial activity against all the bacteria, except *Proteus vulgaris* and its 10 % of 50 µl concentration found to be more effective. The aqueous (100 µl) and

ethanolic (50 µl) extract of *Cuminum cyminum* L. found effective against *Proteus vulgaris* only. Ethanolic extract from turmeric shown zone of inhibition against *Bacillus subtilis* only and its 50 µl of 10 % concentration found effective. Aqueous extract of *Piper nigrum* L. doesn't show any inhibition zone against any test organisms but its ethanolic extract shown effective inhibition zone against all the bacteria except *Proteus vulgaris*. Only the ethanolic extract of *Curcuma longa* L. was found effective against *Bacillus subtilis* and *Salmonella typhimurium*

and its 50 µl of 10 % concentration was most effective. The results obtained may support the use of the mentioned spices in traditional medicine for the treatment of Rheumatism, Nausea, Diarrhea, Diuretic diseases, digestive system related diseases, in Flatulense, muscle spasma, Anti-inflammatory, in some autoimmune diseases, etc. and in drug developments.

CONCLUSION

The research concludes that present Indian spices are the rich source of valuable secondary metabolites and shown presence of significant antimicrobial properties. Specifically ethanolic extract of *Curcuma longa*, aqueous extract of *Cuminum cyminum*, *Cinnamomum tamala* and *Piper nigrum* yielded more phytochemicals while both solvents found effective for *Rhizoma zingiberis*. Ethanolic extract of *Piper nigrum* and *Rhizoma zingiberis* found most effective antibacterial among all.

Conflict of interests:

The authors declare that there is no conflict of interests regarding the publication of this paper.

Table 5: Antimicrobial activity of Aqueous (A) and Ethanolic (E) extract of *Cinnamomum tamala*

Organism used	Zone of inhibition (in mm)					
	<i>Cinnamomum tamala</i> (10 %)					
	A		E		Control	
	50µl	100 µl	50µl	100 µl	50µl	100 µl
<i>Bacillus subtilis</i> NCIM 2635	-	-	18.0± 1.00	20.6 ±0.5 7	-	-
<i>Proteus vulgaris</i> NCIM 2813	-	-	-	-	-	-
<i>Salmonella typhimurium</i> m NCIM 2501	-	-	17.0± 1.00	14.0 ±1.0 0	-	-
<i>Staphylococcus aureus</i> NCIM 2654	-	-	-	-	-	-

References:

- Patil R. S., Godghate A. G. and Sawant R. S. (2014). Phytochemicals and Antimicrobial activity of leaves of *Homonoia riparia* L. *Int. J. Pharm. Bio. Sci*, 5(2): 352-356.

- Nanasombat, S., Prasertsin, V., Graisin, K., Shain, H. and Thanaboripat, B (2002). Efficacy of New Enzyme-Linked Immunosorbent Assay for Rapid Detection of *Salmonella* in Foods. Government Pharmaceutical Organization Report, Bangkok, 51: 53-57.
- Praveen, T. and Nazia, M.A.C. (2006). Bactericidal Activity of Black Pepper, Aniseed and Coriander Against Oral Isolates. *Pakistan J. Pharmaceutical Sciences*, 19(3): 214-218.
- Indu, M. N., Hatha, A. A. M. and Abirosh, C. (2006). Antimicrobial Activity of Some of the South-Indian Spices Against Serotypes of *Escherichia Coli*. *Brazilian Journal of Microbiology*. 37:153-158.
- Godghate, A. G. and Sawant, R. S. (2014). Secondary Metabolites Determinations Qualitatively from Bark of *Butea monosperma* and *Eucalyptus globules*. *International Journal of Science, Environment and Technology*, 3(2): 497-501.
- Damodaran Ashokan and Manohar Sandhya (2012). *Herbal Tech Industry*, 11-13.
- Patil R. S. and Bhise K. K. (2015). Evaluation of phytochemicals and *in vitro* antimicrobial activity of aqueous and ethanolic extract from seeds of *Ricinus communis* Linn. *European Journal of Biotechnology and Bioscience*, 3(3):19-23.
- Harborne J. B. (1973). *Phytochemical Methods: A guide to modern techniques of plant analysis*. Chapman and Hall. New York, pp. 279. 3rd Edn.
- Sawant R. S. and Godghate A. G. (2013). Qualitative Phytochemical Screening of Rhizomes of *Curcuma Longa* Linn. *International Journal of Science, Environment and Technology*, (2)4: 634 – 641.
- Patil R. S., Harale P. M., Shivangekar K. V., Kumbhar P. P. and Desai R. R. (2015). Phytochemical potential and *in vitro* antimicrobial activity of *Piper betle* Linn. leaf extracts *Journal of Chemical and Pharmaceutical Research*, 7(5):1095-1101.
- Sofowora A. (1993). *Medicinal Plants and Traditional Medicinal in Africa*. 2nd Ed. Sunshine House, Ibadan, Nigeria: Spectrum Book Ltd; pp. 134-156.
- Patil R. S., Desai A. B. and Wagh S. A. (2015). Comparative Study of Antimicrobial Compounds Extracted From Leaves of *Nicotiana Tabacum* and Cigarette. *World Journal of Pharmacy and Pharmaceutical Sciences*, 4(3): 1511-1518.
- Baez D. A., Vallejo L.G.Z. and Jimenez-Estrada M. (1999). Phytochemical studies on *Senna skinneri* and *Senna wishizeni*. *Nat. Prod. Lett.*, Berks, 13: 223-228.
- Ogundipe O. O., Moody J. O., Houghton P. J. and Odelola H. A. (2001). Bioactive chemical

- constituents from *Alchornea laxiflora* (benth) pax and hoffman. *J. Ethnopharmacol.*, Lausanne, 74: 275-280.
15. Xu HX and Lee S. F. (2001). Activity of plant flavonoids against antibiotic-resistant bacteria. *Phytother. Res.*, London, 15: 39-43.
 16. Harborne J. B. and Willians C. A. (2000). Advances in flavonoid research since 1992. *Phytochemistry*, Oxford, 55: 481-504.
 17. Lin LU, Shu-wen L, Shi-bo J and Shu-guang W. (2004). Tannin inhibits HIV-1 entry by targeting gp 41. *Acta Pharmacol Sin.* 25(2): 213-218.
 18. Akiyama H, Kazuyasu F, Yamasaki O, Oono T and Iwatsuki K. (2001). Antibacterial action of several tannins against *Staphylococcus aureus*. *J. Antimicrobial Chemotherapy.* 48(48): 487-491.
 19. Funatogawa K, Hayashi S, Shimomura H, Yoshida T, Hatano T, Ito H and Iría Y (2004). Antibacterial activity of hydrolysable tannins derived from medicinal plants against *Helicobacter pylori*. *Microbiol. Immunol.* 48(4): 251-261.
 20. Venkanna Lunavath and Estari Mamidala (2013). Preliminary phytochemical screening and antibacterial studies of the leaves of *Eclipta alba* (L). *Int J Pharm Bio Sci* 2013 July; 4(3): (B) 380 – 384
 21. G. Krishna, M.A Singara Charya and V. Praniitha. 2014. Evaluation of antibacterial and antifungal activity of fruiting body extracts of *Trametes versicolor*. *Biolife.* 2(4). 1181-1184
